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Molecular MetaGraph and Molecular Iterated MetaGraph in Chemistry and BioChemistry

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Abstract

Graph theory investigates mathematical structures consisting of vertices and edges, providing a foundation for modeling relationships and connectivity. A *MetaGraph* is a higher-level graph in which the vertices are themselves graphs, with edges representing specified relations among these graphs. An *Iterated MetaGraph* extends this concept recursively: its vertices are MetaGraphs, thereby forming a hierarchy of graph-of-graphs structures across multiple levels. In this paper, we introduce two new extensions, the *Molecular MetaGraph* and the *Molecular Iterated MetaGraph*, which generalize the concept of molecular graphs through the frameworks of MetaGraphs and Iterated MetaGraphs. Furthermore, we provide illustrative applications of these models in biochemistry. These applications demonstrate how an iterative, meta-level perspective can be applied to molecular graphs, offering new insights into biochemical structures and processes.

1 Preliminaries

This section presents the fundamental concepts and definitions that underpin the discussions in this paper. Unless otherwise noted, all graphs considered here are *finite*.

1.1 MetaGraph(Graph of Graph)

Graph theory investigates mathematical structures consisting of vertices and edges to model relationships and connectivity [1,2]. A MetaGraph is a graph whose vertices are themselves graphs, with edges representing specified relations between those graphs (cf. [3–5]).

Definition 1.1 (Metagraph (graph of graphs)). (cf. [6]) Fix a nonempty universe \mathfrak{G} of finite graphs (undirected, loopless by default) and a nonempty family of binary relations

$$\mathcal{R} \subseteq \mathcal{P}(\mathfrak{G} \times \mathfrak{G}).$$

A *metagraph over* $(\mathfrak{G}, \mathcal{R})$ is a directed, labelled multigraph

$$M = (V, E, s, t, \lambda)$$

with

$$V \subseteq \mathfrak{G}, \quad s, t : E \rightarrow V, \quad \lambda : E \rightarrow \mathcal{R},$$

satisfying the incidence constraint

$$\forall e \in E : (s(e), t(e)) \in \lambda(e).$$

Elements of V are *meta-vertices* (each is a graph $G \in \mathfrak{G}$). For $e \in E$ with $\lambda(e) = R$, we write $s(e) \xrightarrow{R} t(e)$ and call e a *meta-edge*. If $\mathcal{R} = \{R\}$ is a singleton, labels may be omitted. If every $R \in \mathcal{R}$ is symmetric, M can be viewed as an undirected labelled multigraph.

Remark 1.2. We fix a common base universe \mathfrak{G} of finite, loopless graphs that encode biochemical modules. Each $G \in \mathfrak{G}$ comes with a vertex-labeling $\ell_V : V(G) \rightarrow \Sigma_V$ (e.g. metabolite/protein/gene names) and, when needed, an edge-labeling $\ell_E : E(G) \rightarrow \Sigma_E$ (e.g. reaction/interaction types). We use the following chemically meaningful binary relations on \mathfrak{G} :

$$\begin{aligned} R_{\text{shareM}}(G, H) &: \iff \{\text{metabolite names in } G\} \cap \{\text{metabolite names in } H\} \neq \emptyset, \\ R_{\text{shareP}}(G, H) &: \iff \{\text{protein names in } G\} \cap \{\text{protein names in } H\} \neq \emptyset, \\ R_{\text{GRN}\Delta}(G, H) &: \iff E(G) \Delta E(H) \neq \emptyset \quad (\text{directed GRNs; activation/inhibition labels allowed}). \end{aligned}$$

Set $\mathcal{R} := \{R_{\text{shareM}}, R_{\text{shareP}}, R_{\text{GRN}\Delta}\}$.

Each example specifies a metagraph $M = (V, E, s, t, \lambda)$ over $(\mathfrak{G}, \mathcal{R})$ and verifies the incidence constraint.

Example 1.3 (Metabolic module metagraph: glycolysis–link–TCA). Let the pathway graphs be

$$\begin{aligned} G_{\text{Gly}} : V &= \{\text{Glc, G6P, F6P, Pyr}\}, E = \{\text{Glc} \rightarrow \text{G6P}, \text{G6P} \rightarrow \text{F6P}, \text{F6P} \rightarrow \text{Pyr}\}, \\ G_{\text{PDH}} : V &= \{\text{Pyr, AcCoA, CO}_2\}, E = \{\text{Pyr} \rightarrow \text{AcCoA}\}, \\ G_{\text{TCA}} : V &= \{\text{AcCoA, OAA, Citrate, Succinate, Malate}\}, E = \{\text{AcCoA} + \text{OAA} \rightarrow \text{Citrate}, \dots\}. \end{aligned}$$

Define the metagraph M_{metab} by

$$\begin{aligned} V &= \{G_{\text{Gly}}, G_{\text{PDH}}, G_{\text{TCA}}\}, \quad E = \{e_1, e_2\}, \\ s(e_1) &= G_{\text{Gly}}, \quad t(e_1) = G_{\text{PDH}}, \quad \lambda(e_1) = R_{\text{shareM}} \quad (\text{witness Pyr}), \\ s(e_2) &= G_{\text{PDH}}, \quad t(e_2) = G_{\text{TCA}}, \quad \lambda(e_2) = R_{\text{shareM}} \quad (\text{witness AcCoA}). \end{aligned}$$

For each $e \in E$, $(s(e), t(e)) \in \lambda(e)$ by the indicated shared metabolite, so the incidence constraint holds.

Example 1.4 (Protein-complex metagraph: shared subunits). Let simple complex graphs be

$$\begin{aligned} G_{\text{CplxA}} : V &= \{E_1, E_2\}, E = \{\{E_1, E_2\}\}, \\ G_{\text{CplxB}} : V &= \{E_2, E_3\}, E = \{\{E_2, E_3\}\}, \\ G_{\text{CplxC}} : V &= \{E_4\}, E = \emptyset. \end{aligned}$$

Define the metagraph M_{cplx} by

$$V = \{G_{\text{CplxA}}, G_{\text{CplxB}}, G_{\text{CplxC}}\}, \quad E = \{e_{AB}\}, \quad s(e_{AB}) = G_{\text{CplxA}}, \quad t(e_{AB}) = G_{\text{CplxB}}, \quad \lambda(e_{AB}) = R_{\text{shareP}},$$

witnessed by the shared subunit E_2 . Then $(s(e_{AB}), t(e_{AB})) \in R_{\text{shareP}}$; no edge is drawn to G_{CplxC} because E_{CplxC} shares no protein.

Example 1.5 (Condition-specific GRN metagraph: differential regulation). Let directed GRNs (edges labeled by $\{\text{act}, \text{inh}\}$) be

$$\begin{aligned} G_{\text{GRN}}^{(A)} : V &= \{\text{TF1}, \text{G1}, \text{G2}\}, E = \{\text{TF1} \xrightarrow{\text{act}} \text{G1}, \text{TF1} \xrightarrow{\text{act}} \text{G2}\}, \\ G_{\text{GRN}}^{(B)} : V &= \{\text{TF1}, \text{G1}, \text{G2}\}, E = \{\text{TF1} \xrightarrow{\text{act}} \text{G1}\}, \\ G_{\text{GRN}}^{(C)} : V &= \{\text{TF2}, \text{G2}\}, E = \{\text{TF2} \xrightarrow{\text{inh}} \text{G2}\}. \end{aligned}$$

Define M_{grn} by

$$\begin{aligned} V &= \{G_{\text{GRN}}^{(A)}, G_{\text{GRN}}^{(B)}, G_{\text{GRN}}^{(C)}\}, \quad E = \{e_{AB}, e_{AC}\}, \\ s(e_{AB}) &= G_{\text{GRN}}^{(A)}, \quad t(e_{AB}) = G_{\text{GRN}}^{(B)}, \quad \lambda(e_{AB}) = R_{\text{GRN}\Delta} \quad (\text{witness: edge } \text{TF1} \rightarrow \text{G2} \text{ present only in } A), \\ s(e_{AC}) &= G_{\text{GRN}}^{(A)}, \quad t(e_{AC}) = G_{\text{GRN}}^{(C)}, \quad \lambda(e_{AC}) = R_{\text{GRN}\Delta} \quad (\text{many-edge difference}). \end{aligned}$$

Thus $(s(e), t(e)) \in \lambda(e)$ for each e , so M_{grn} is a valid metagraph.

1.2 Iterated MetaGraph(Graph of Graph of ... of Graph)

An Iterated MetaGraph is a graph whose vertices are metagraphs, recursively extending graph-of-graphs structure to multiple hierarchical levels [7].

Definition 1.6 (Unit metagraph embedding). [7] For $X \in \mathfrak{G}$ define the *unit metagraph*

$$U(X) := (\{X\}, \emptyset, _, _, _).$$

This gives an injective map $U : \mathfrak{G} \hookrightarrow \text{Obj}(\text{Meta}(\mathfrak{G}, \mathcal{R}))$.

Definition 1.7 (Relation lifting). Given \mathcal{R} on \mathfrak{G} , define its *lift* \mathcal{R}^\uparrow on finite metagraphs over $(\mathfrak{G}, \mathcal{R})$ by

$$\forall R \in \mathcal{R}, \quad (M_1, M_2) \in R^\uparrow \iff \exists x \in V(M_1), y \in V(M_2) : (x, y) \in R.$$

Set $\mathcal{R}^\uparrow := \{R^\uparrow : R \in \mathcal{R}\}$.

Definition 1.8 (Iterated object and relation universes). Define recursively for $t \in \mathbb{N}_0$:

$$\begin{aligned} \mathfrak{G}^{(0)} &:= \mathfrak{G}, \quad \mathcal{R}^{(0)} := \mathcal{R}, \\ \mathfrak{G}^{(t+1)} &:= \left\{ \text{finite metagraphs over } (\mathfrak{G}^{(t)}, \mathcal{R}^{(t)}) \right\}, \quad \mathcal{R}^{(t+1)} := (\mathcal{R}^{(t)})^\uparrow. \end{aligned}$$

Definition 1.9 (Iterated MetaGraph of depth t). For $t \in \mathbb{N}_0$, an *iterated metagraph of depth t* is a metagraph

$$M^{(t)} = (V^{(t)}, E^{(t)}, s^{(t)}, t^{(t)}, \lambda^{(t)})$$

over $(\mathfrak{G}^{(t)}, \mathcal{R}^{(t)})$, i.e., $V^{(t)} \subseteq \mathfrak{G}^{(t)}$, $\lambda^{(t)} : E^{(t)} \rightarrow \mathcal{R}^{(t)}$ and

$$\forall e \in E^{(t)} : (s^{(t)}(e), t^{(t)}(e)) \in \lambda^{(t)}(e).$$

Remark 1.10. We use the relation lifting of the preliminaries: for $R \in \mathcal{R}$ and metagraphs M_1, M_2 over $(\mathfrak{G}, \mathcal{R})$,

$$(M_1, M_2) \in R^\uparrow \iff \exists X \in V(M_1), \exists Y \in V(M_2) \text{ such that } (X, Y) \in R.$$

Example 1.11 (Depth $t = 1$: Metabolism vs. complexes, linked by an annotation map). Let M_{metab} and M_{cplx} be as above. Define a base cross-relation $R_{\text{annot}} \subseteq \mathfrak{G} \times \mathfrak{G}$ by

$$(G, H) \in R_{\text{annot}} \iff \exists \text{enzyme } E \in V(H) \text{ that catalyzes some edge of } G.$$

(Equivalently, fix an annotation map $\kappa : \{\text{enzymes}\} \rightarrow \{\text{reactions}\}$ and require $\kappa(E) \in E(G)$.) Assume $\kappa(E_1) = \text{G6P} \rightarrow \text{F6P}$ (a step in G_{Gly}) and $E_1 \in V(G_{\text{CplxA}})$. Then $(G_{\text{Gly}}, G_{\text{CplxA}}) \in R_{\text{annot}}$. Form the depth-1 metagraph

$$N_1^{(1)} = (\{M_{\text{metab}}, M_{\text{cplx}}\}, \{f\}, s^{(1)}, t^{(1)}, \lambda^{(1)}),$$

with $s^{(1)}(f) = M_{\text{metab}}$, $t^{(1)}(f) = M_{\text{cplx}}$, and $\lambda^{(1)}(f) = R_{\text{annot}}^\uparrow$. By the witness $(G_{\text{Gly}}, G_{\text{CplxA}})$, $(s^{(1)}(f), t^{(1)}(f)) \in \lambda^{(1)}(f)$, so $N_1^{(1)}$ is valid.

Example 1.12 (Depth $t = 1$: Comparative metabolism across species). Build two organism-specific metagraphs using R_{shareM} :

$$M_{\text{met}}^{\text{Ec}} : V = \{G_{\text{Gly}}^{\text{Ec}}, G_{\text{TCA}}^{\text{Ec}}\}, E = \{G_{\text{Gly}}^{\text{Ec}} \xrightarrow{R_{\text{shareM}}} G_{\text{TCA}}^{\text{Ec}}\}$$

with witness metabolite AcCoA (via the explicit PDH-link included in $G_{\text{Gly}}^{\text{Ec}}$), and

$$M_{\text{met}}^{\text{Sc}} : V = \{G_{\text{Gly}}^{\text{Sc}}, G_{\text{TCA}}^{\text{Sc}}\}, E = \{G_{\text{Gly}}^{\text{Sc}} \xrightarrow{R_{\text{shareM}}} G_{\text{TCA}}^{\text{Sc}}\}$$

with the same witness. At depth 1, set

$$N_2^{(1)} = (\{M_{\text{met}}^{\text{Ec}}, M_{\text{met}}^{\text{Sc}}\}, \{g\}, s^{(1)}, t^{(1)}, \lambda^{(1)}), \quad s^{(1)}(g) = M_{\text{met}}^{\text{Ec}}, \quad t^{(1)}(g) = M_{\text{met}}^{\text{Sc}}, \quad \lambda^{(1)}(g) = R_{\text{shareM}}^\uparrow.$$

Since $G_{\text{TCA}}^{\text{Ec}}$ and $G_{\text{TCA}}^{\text{Sc}}$ both contain the metabolite *Citrate*, $(G_{\text{TCA}}^{\text{Ec}}, G_{\text{TCA}}^{\text{Sc}}) \in R_{\text{shareM}}$, which witnesses $(s^{(1)}(g), t^{(1)}(g)) \in \lambda^{(1)}(g)$.

Example 1.13 (Depth $t = 2$: Linking depth-1 projects by double lift). Let $N_1^{(1)}$ and $N_2^{(1)}$ be as above. Consider the depth-2 metagraph

$$Q^{(2)} = (V^{(2)}, E^{(2)}, s^{(2)}, t^{(2)}, \lambda^{(2)}), \quad V^{(2)} = \{N_1^{(1)}, N_2^{(1)}\}, \quad E^{(2)} = \{h\}.$$

Define

$$s^{(2)}(h) = N_1^{(1)}, \quad t^{(2)}(h) = N_2^{(1)}, \quad \lambda^{(2)}(h) = (R_{\text{shareM}}^\uparrow)^\uparrow.$$

To witness $(s^{(2)}(h), t^{(2)}(h)) \in \lambda^{(2)}(h)$, pick $X = M_{\text{metab}} \in V(N_1^{(1)})$ and $Y = M_{\text{met}}^{\text{Ec}} \in V(N_2^{(1)})$. Since $G_{\text{TCA}} \in V(X)$ and $G_{\text{TCA}}^{\text{Ec}} \in V(Y)$ share metabolite *Citrate*, we have $(G_{\text{TCA}}, G_{\text{TCA}}^{\text{Ec}}) \in R_{\text{shareM}}$, hence $(X, Y) \in R_{\text{shareM}}^\uparrow$, which by definition yields $(s^{(2)}(h), t^{(2)}(h)) \in (R_{\text{shareM}}^\uparrow)^\uparrow$. Thus $Q^{(2)}$ is a valid depth-2 iterated metagraph.

1.3 Molecular Graph

A *Molecular Graph* models a molecule with atoms as vertices and bonds as edges, thereby representing its structural connectivity [8–10]. Related concepts include the *Molecular HyperGraph* [11,12] and the *Molecular SuperHyperGraph* [13, 14], which provide generalized frameworks for capturing higher-order interactions in molecular structures. Also the related concepts such as Chemical Graphs [15, 16], Neutrosophic Chemical Graphs [17], and Molecular Fuzzy Graphs [18] are also well known.

Definition 1.14 (Molecular Graph). (cf. [19, 20]) A *molecular graph* is a finite, simple, undirected graph $G = (V, E)$ in which each vertex $v \in V$ represents an atom and each edge $e = \{u, v\} \in E$ represents a chemical bond between atoms u and v . (Optionally, vertex/edge labels may encode atom types and bond types or orders.)

Example 1.15 (Water H_2O as a molecular graph). Let

$$V = \{O, H_1, H_2\}, \quad E = \{\{O, H_1\}, \{O, H_2\}\}.$$

Then $G = (V, E)$ is a finite, simple, undirected molecular graph: each vertex represents an atom (oxygen O , hydrogens H_1, H_2) and each edge represents an O–H bond. Optionally, one may add labels

$$\tau(O) = O, \quad \tau(H_i) = H \ (i = 1, 2), \quad \beta(\{O, H_i\}) = \text{single},$$

and the adjacency matrix (ordering O, H_1, H_2) is

$$A = \begin{pmatrix} 0 & 1 & 1 \\ 1 & 0 & 0 \\ 1 & 0 & 0 \end{pmatrix}, \quad \deg(O) = 2, \quad \deg(H_1) = \deg(H_2) = 1.$$

2 Main Results of this Paper

This section presents the main results established in this paper.

2.1 Molecular MetaGraph

A Molecular MetaGraph is a higher-level graph where vertices are molecular graphs and edges encode chemically invariant relations among them. We first fix a typed setting for molecules.

Definition 2.1 (Typed molecular graph and type-preserving isomorphism). Let Σ_V be a finite set of *atom types* (e.g. element symbols, hybridizations, charges) and Σ_E a finite set of *bond types* (e.g. single, double, triple, aromatic). A *typed molecular graph* is a quadruple

$$G = (V, E, \tau, \beta),$$

where V is a finite vertex set (atoms), $E \subseteq \binom{V}{2}$ (bonds), $\tau : V \rightarrow \Sigma_V$ assigns each atom its type, and $\beta : E \rightarrow \Sigma_E$ assigns each bond its type. A bijection $\varphi : V(G) \rightarrow V(H)$ between typed molecular graphs G and H is a *type-preserving isomorphism* (notation $G \cong H$) if

$$\{u, v\} \in E(G) \iff \{\varphi(u), \varphi(v)\} \in E(H), \quad \tau_H(\varphi(v)) = \tau_G(v), \quad \beta_H(\{\varphi(u), \varphi(v)\}) = \beta_G(\{u, v\}).$$

Let \mathfrak{Mol} denote the universe of all finite typed molecular graphs (Definition 2.1).

Definition 2.2 (Chemically invariant relations). A binary relation $R \subseteq \mathfrak{Mol} \times \mathfrak{Mol}$ is *chemically invariant* if for all $G, H, G', H' \in \mathfrak{Mol}$,

$$G \cong G' \text{ and } H \cong H' \text{ and } (G, H) \in R \implies (G', H') \in R.$$

Let $\mathcal{R}_{\text{chem}}$ be any nonempty family of chemically invariant relations. Typical examples (not required for the theory) include: substructure (R_{sub}), reaction-step (R_{rxn}), isomer-of (R_{iso}), scaffold-similarity-at-threshold- θ ($R_{\text{sim},\theta}$), etc.

Definition 2.3 (Molecular MetaGraph). A *Molecular MetaGraph over* $(\mathfrak{Mol}, \mathcal{R}_{\text{chem}})$ is a directed, labelled multigraph

$$\mathbf{M} = (V, E, s, t, \lambda)$$

with

$$V \subseteq \mathfrak{Mol}, \quad s, t : E \rightarrow V, \quad \lambda : E \rightarrow \mathcal{R}_{\text{chem}},$$

satisfying the metagraph incidence constraint

$$\forall e \in E : (s(e), t(e)) \in \lambda(e).$$

The elements of V are *meta-vertices* (each is a molecular graph), and each $e \in E$ is a *meta-edge* labelled by a chemical relation $\lambda(e)$ that holds on its endpoints.

Remark 2.4. We use the chemically invariant base relations

$$R_{\text{sub}}, R_{\text{rxn}}, R_{\text{iso}}, R_{\text{sim},\theta}, R_{\text{eq}} \in \mathcal{R}_{\text{chem}},$$

where R_{sub} denotes “is a (typed) substructure of,” R_{rxn} denotes “reactant-to-product in a fixed elementary step,” R_{iso} denotes “constitutional isomers,” $R_{\text{sim},\theta}$ denotes “scaffold similarity at threshold θ ,” and R_{eq} is molecular identity (type-preserving isomorphism). Recall that a Molecular MetaGraph is $\mathbf{M} = (V, E, s, t, \lambda)$ with $V \subseteq \mathfrak{Mol}$ and $\forall e \in E : (s(e), t(e)) \in \lambda(e)$.

Example 2.5 (Molecular MetaGraph I: A single-step reaction mini-network (Fischer esterification)). Let the molecular vertices be

$$V = \{G_{\text{AA}}, G_{\text{EtOH}}, G_{\text{EA}}, G_{\text{H}_2\text{O}}\},$$

standing for acetic acid (SMILES CC(=O)O), ethanol (CCO), ethyl acetate (CCOC(=O)C), and water (O). Define four directed meta-edges encoding reactant→product incidence in the esterification step:

$$\begin{aligned} E = \{e_1, e_2, e_3, e_4\}, \quad & s(e_1) = G_{\text{AA}}, t(e_1) = G_{\text{EA}}, \lambda(e_1) = R_{\text{rxn}}, \\ & s(e_2) = G_{\text{EtOH}}, t(e_2) = G_{\text{EA}}, \lambda(e_2) = R_{\text{rxn}}, \\ & s(e_3) = G_{\text{AA}}, t(e_3) = G_{\text{H}_2\text{O}}, \lambda(e_3) = R_{\text{rxn}}, \\ & s(e_4) = G_{\text{EtOH}}, t(e_4) = G_{\text{H}_2\text{O}}, \lambda(e_4) = R_{\text{rxn}}. \end{aligned}$$

Then $\mathbf{M}_{\text{ester}} = (V, E, s, t, \lambda)$ is a Molecular MetaGraph over $(\mathfrak{Mol}, \mathcal{R}_{\text{chem}})$.

Example 2.6 (Molecular MetaGraph II: Substructure & similarity on simple aromatics). Let

$$V = \{G_{\text{Bz}}, G_{\text{Tol}}, G_{\text{Ph}}\},$$

for benzene (c1ccccc1), toluene (Cc1ccccc1), and phenol (Oc1ccccc1). Define

$$E = \{e_1, e_2, e_3\},$$

with

$$\begin{aligned} s(e_1) &= G_{\text{Bz}}, \quad t(e_1) = G_{\text{Tol}}, \quad \lambda(e_1) = R_{\text{sub}}, \\ s(e_2) &= G_{\text{Bz}}, \quad t(e_2) = G_{\text{Ph}}, \quad \lambda(e_2) = R_{\text{sub}}, \\ s(e_3) &= G_{\text{Tol}}, \quad t(e_3) = G_{\text{Ph}}, \quad \lambda(e_3) = R_{\text{sim}, 0.6}. \end{aligned}$$

Interpreting $R_{\text{sim}, 0.6}$ as “Tanimoto (scaffold) similarity ≥ 0.6 ,” $M_{\text{arom}} = (V, E, s, t, \lambda)$ is a Molecular MetaGraph in which benzene is a typed substructure of toluene and phenol, and toluene is scaffold-similar to phenol.

Example 2.7 (Molecular MetaGraph III: Constitutional isomers of $\text{C}_3\text{H}_8\text{O}$). Let

$$V = \{G_{n\text{-PrOH}}, G_{i\text{-PrOH}}, G_{\text{MeOEt}}\},$$

for 1-propanol (CCCC), isopropanol (CC(O)C), and methoxyethane (CCOC). Since R_{iso} is symmetric, we realize it as a bidirected triangle:

$$E = \{e_{12}, e_{21}, e_{13}, e_{31}, e_{23}, e_{32}\},$$

where, writing $(1, 2, 3) = (n\text{-PrOH}, i\text{-PrOH}, \text{MeOEt})$,

$$s(e_{ij}) = G_i, \quad t(e_{ij}) = G_j, \quad \lambda(e_{ij}) = R_{\text{iso}} \quad (i \neq j).$$

Then $M_{\text{isoC3H8O}} = (V, E, s, t, \lambda)$ is a Molecular MetaGraph whose meta-edges encode isomerism.

Theorem 2.8 (Every Molecular MetaGraph is a MetaGraph). *With $\mathfrak{G} := \mathfrak{Mol}$ and $\mathcal{R} := \mathcal{R}_{\text{chem}}$, every structure M as in Definition 2.3 is a metagraph over $(\mathfrak{G}, \mathcal{R})$ in the sense of the preliminaries.*

Proof. Comparing Definition 2.3 with the metagraph schema $M = (V, E, s, t, \lambda)$, we must verify $V \subseteq \mathfrak{G}$, $s, t : E \rightarrow V$, $\lambda : E \rightarrow \mathcal{R}$, and the incidence condition $(s(e), t(e)) \in \lambda(e)$ for all $e \in E$. By construction, $V \subseteq \mathfrak{Mol} = \mathfrak{G}$, s, t, λ have the required codomains, and the last property is explicitly imposed in Definition 2.3. Therefore M is a metagraph over $(\mathfrak{G}, \mathcal{R})$. \square

Next we show that Molecular MetaGraphs *generalize* ordinary molecular graphs by a faithful embedding that preserves all atom and bond information.

Definition 2.9 (Atomic one-vertex templates). For each $a \in \Sigma_V$, let $K_1^{(a)}$ denote the one-vertex graph with vertex-type a and no edges; formally,

$$K_1^{(a)} = (\{\ast\}, \emptyset, \tau_{K_1^{(a)}}(\ast) := a, \beta \text{ undefined}) \in \mathfrak{Mol}.$$

Definition 2.10 (Bond-labelled meta-relations). For each $b \in \Sigma_E$, define a relation $R_{\text{bond}, b} \subseteq \mathfrak{Mol} \times \mathfrak{Mol}$ by

$$(K_1^{(a)}, K_1^{(a')}) \in R_{\text{bond}, b}$$

\iff “there exists a molecular graph containing a b -bond between an a -atom and an a' -atom”.

This relation is chemically invariant (it depends only on types), hence $R_{\text{bond}, b} \in \mathcal{R}_{\text{chem}}$ after enlarging $\mathcal{R}_{\text{chem}}$ if necessary.

Definition 2.11 (Canonical embedding $J : \mathfrak{Mol} \hookrightarrow \text{MolMeta}$). Given $G = (V, E, \tau, \beta) \in \mathfrak{Mol}$, define a Molecular MetaGraph

$$J(G) := (V^*, E^*, s^*, t^*, \lambda^*)$$

as follows:

$$\begin{aligned} V^* &:= \{ K_1^{(\tau(v))} \mid v \in V \}, \\ E^* &:= \{ e_{uv}^* \mid \{u, v\} \in E \} \text{ (a multiset if needed),} \\ s^*(e_{uv}^*) &:= K_1^{(\tau(u))}, \quad t^*(e_{uv}^*) := K_1^{(\tau(v))}, \\ \lambda^*(e_{uv}^*) &:= R_{\text{bond}, \beta(\{u, v\})}. \end{aligned}$$

Let $\pi : V^* \rightarrow V$ be the bijection $\pi(K_1^{(\tau(v))}) := v$.

Theorem 2.12 (Molecular MetaGraphs generalize molecular graphs). *For every typed molecular graph $G = (V, E, \tau, \beta)$, the “collapse” of $J(G)$ recovers G exactly:*

$$\text{col}(J(G)) = G,$$

where

$$\text{col}(J(G)) := (V, E, \tau, \beta)$$

with V and τ, β taken from G and E reconstructed from $J(G)$ by

$$E = \left\{ \{ \pi(s^*(e^*)), \pi(t^*(e^*)) \} : e^* \in E^*, \lambda^*(e^*) = R_{\text{bond}, \beta(\{\pi(s^*(e^*)), \pi(t^*(e^*))\})} \right\}.$$

Consequently, J is injective on isomorphism classes and embeds the category of typed molecular graphs (with type-preserving isomorphisms) fully and faithfully into the category of Molecular MetaGraphs whose edges use only $\{R_{\text{bond}, b} : b \in \Sigma_E\}$.

Proof. Fix $G = (V, E, \tau, \beta)$ and build $J(G)$ as in Definition 2.11. By construction, $\pi : V^* \rightarrow V$ is a bijection. We verify equality of edge sets and labels by direct elementwise computation.

(\subseteq) Let $e^* = e_{uv}^* \in E^*$. Then by definition,

$$s^*(e^*) = K_1^{(\tau(u))}, \quad t^*(e^*) = K_1^{(\tau(v))}, \quad \lambda^*(e^*) = R_{\text{bond}, \beta(\{u, v\})}.$$

Applying π we obtain

$$\{\pi(s^*(e^*)), \pi(t^*(e^*))\} = \{u, v\} \in E,$$

and the recovered bond type is

$$\beta(\{\pi(s^*(e^*)), \pi(t^*(e^*))\}) = \beta(\{u, v\}),$$

which matches the label index of $\lambda^*(e^*)$. Hence every meta-edge contributes exactly the original bond.

(\supseteq) Conversely, let $\{u, v\} \in E$. Then $e_{uv}^* \in E^*$ by construction, and

$$\{\pi(s^*(e_{uv}^*)), \pi(t^*(e_{uv}^*))\} = \{u, v\}, \quad \lambda^*(e_{uv}^*) = R_{\text{bond}, \beta(\{u, v\})},$$

so the edge $\{u, v\}$ is recovered in $\text{col}(J(G))$ with the correct bond type.

Thus the recovered edge multiset and labels coincide elementwise with those of G , and vertex types are preserved by π :

$$\tau(\pi(K_1^{(\tau(v))})) = \tau(v).$$

Therefore $\text{col}(J(G)) = G$ as graphs with types. This proves that J is injective on objects up to isomorphism and, by the explicit reconstruction, that hom-sets are preserved (faithfulness). \square

2.2 Molecular Iterated MetaGraph

A Molecular Iterated MetaGraph recursively treats Molecular MetaGraphs as vertices, building multi-level hierarchies of graph-of-graphs structures across chemistry. We lift chemically invariant relations level by level, as in the iterated metagraph construction from the preliminaries, but restricted to \mathfrak{Mol} and $\mathcal{R}_{\text{chem}}$.

Definition 2.13 (Lifted chemical relations). Set $\mathfrak{Mol}^{(0)} := \mathfrak{Mol}$ and $\mathcal{R}_{\text{chem}}^{(0)} := \mathcal{R}_{\text{chem}}$. Given $\mathfrak{Mol}^{(t)}$ and $\mathcal{R}_{\text{chem}}^{(t)}$, define

$$\mathfrak{Mol}^{(t+1)} := \left\{ \text{finite Molecular MetaGraphs over } (\mathfrak{Mol}^{(t)}, \mathcal{R}_{\text{chem}}^{(t)}) \right\},$$

and for each $R \in \mathcal{R}_{\text{chem}}^{(t)}$ its lift $R^\uparrow \subseteq \mathfrak{Mol}^{(t+1)} \times \mathfrak{Mol}^{(t+1)}$ by

$$(\mathsf{M}_1, \mathsf{M}_2) \in R^\uparrow \iff \exists X \in V(\mathsf{M}_1), Y \in V(\mathsf{M}_2) \text{ with } (X, Y) \in R.$$

Set $\mathcal{R}_{\text{chem}}^{(t+1)} := \{R^\uparrow : R \in \mathcal{R}_{\text{chem}}^{(t)}\}$.

Definition 2.14 (Molecular Iterated MetaGraph of depth t). For $t \in \mathbb{N}_0$, a *Molecular Iterated MetaGraph of depth t* is a metagraph

$$\mathsf{M}^{(t)} = (V^{(t)}, E^{(t)}, s^{(t)}, t^{(t)}, \lambda^{(t)})$$

over $(\mathfrak{Mol}^{(t)}, \mathcal{R}_{\text{chem}}^{(t)})$, i.e.,

$$V^{(t)} \subseteq \mathfrak{Mol}^{(t)}, \quad \lambda^{(t)} : E^{(t)} \rightarrow \mathcal{R}_{\text{chem}}^{(t)}, \quad \forall e \in E^{(t)} : (s^{(t)}(e), t^{(t)}(e)) \in \lambda^{(t)}(e).$$

Remark 2.15. For iterated examples we use the lifted relations of Definition 2.14: for $R \in \mathcal{R}_{\text{chem}}^{(t)}$, its lift R^\uparrow satisfies

$$(\mathsf{M}_1, \mathsf{M}_2) \in R^\uparrow \iff \exists X \in V(\mathsf{M}_1), \exists Y \in V(\mathsf{M}_2) \text{ with } (X, Y) \in R.$$

Example 2.16 (Molecular Iterated MetaGraph I (depth $t = 1$): Substructure lifted across two meta-graphs).

Let $\mathsf{M}_A = \mathsf{M}_{\text{arom}}$ from Example 2 restricted to $V_A = \{G_{\text{Bz}}, G_{\text{Tol}}\}$ and edge $G_{\text{Bz}} \xrightarrow{R_{\text{sub}}} G_{\text{Tol}}$. Let M_B have $V_B = \{G_{\text{Bz}}, G_{\text{Ph}}\}$ and edge $G_{\text{Bz}} \xrightarrow{R_{\text{sub}}} G_{\text{Ph}}$. Define a depth-1 iterated meta-graph

$$\mathsf{N}^{(1)} = (V^{(1)}, E^{(1)}, s^{(1)}, t^{(1)}, \lambda^{(1)}),$$

with

$$V^{(1)} = \{\mathsf{M}_A, \mathsf{M}_B\}, \quad E^{(1)} = \{f\}, \quad s^{(1)}(f) = \mathsf{M}_A, \quad t^{(1)}(f) = \mathsf{M}_B, \quad \lambda^{(1)}(f) = R_{\text{sub}}^\uparrow.$$

Since $(G_{\text{Bz}}, G_{\text{Ph}}) \in R_{\text{sub}}$ with $G_{\text{Bz}} \in V(\mathsf{M}_A)$ and $G_{\text{Ph}} \in V(\mathsf{M}_B)$, we have $f \in R_{\text{sub}}^\uparrow$ by definition, so $\mathsf{N}^{(1)}$ is a valid Molecular Iterated MetaGraph of depth 1.

Example 2.17 (Molecular Iterated MetaGraph II (depth $t = 1$): Equality-lift between forward and reverse steps). Let $\mathsf{M}_{\text{ester}}$ be Example 1 (forward esterification). Let M_{hydr} be the corresponding hydrolysis meta-graph with edges $G_{\text{EA}} \rightarrow G_{\text{AA}}, G_{\text{EA}} \rightarrow G_{\text{EtOH}}, G_{\text{H}_2\text{O}} \rightarrow G_{\text{AA}}, G_{\text{H}_2\text{O}} \rightarrow G_{\text{EtOH}}$ all labelled R_{rxn} . Define

$$\mathsf{H}^{(1)} = (\{\mathsf{M}_{\text{ester}}, \mathsf{M}_{\text{hydr}}\}, \{g_1, g_2\}, s^{(1)}, t^{(1)}, \lambda^{(1)}),$$

with

$$s^{(1)}(g_1) = \mathsf{M}_{\text{ester}}, \quad t^{(1)}(g_1) = \mathsf{M}_{\text{hydr}}, \quad s^{(1)}(g_2) = \mathsf{M}_{\text{hydr}}, \quad t^{(1)}(g_2) = \mathsf{M}_{\text{ester}}, \quad \lambda^{(1)}(g_1) = \lambda^{(1)}(g_2) = R_{\text{eq}}^\uparrow.$$

Because both meta-graphs contain G_{EA} (and other common molecules), there exist $X = Y = G_{\text{EA}}$ with $(X, Y) \in R_{\text{eq}}$; hence $(\mathsf{M}_{\text{ester}}, \mathsf{M}_{\text{hydr}}) \in R_{\text{eq}}^\uparrow$ and $\mathsf{H}^{(1)}$ is a valid depth-1 Molecular Iterated MetaGraph.

Example 2.18 (Molecular Iterated MetaGraph III (depth $t = 2$): Project-level linkage via double lift). Form two depth-1 nodes:

$$\mathsf{P}_1^{(1)} := (\{\mathsf{M}_A, \mathsf{M}_{\text{ester}}\}, \{p\}, s^{(1)}, t^{(1)}, \lambda^{(1)}), \quad s^{(1)}(p) = \mathsf{M}_A, \quad t^{(1)}(p) = \mathsf{M}_{\text{ester}}, \quad \lambda^{(1)}(p) = R_{\text{eq}}^\uparrow,$$

witnessed by the shared molecule $G_{\text{H}_2\text{O}}$ (or any chosen common species), and

$$\mathsf{P}_2^{(1)} := (\{\mathsf{M}_B, \mathsf{M}_{\text{hydr}}\}, \{q\}, s^{(1)}, t^{(1)}, \lambda^{(1)}), \quad s^{(1)}(q) = \mathsf{M}_B, \quad t^{(1)}(q) = \mathsf{M}_{\text{hydr}}, \quad \lambda^{(1)}(q) = R_{\text{eq}}^\uparrow,$$

witnessed by G_{AA} or G_{EtOH} . Now define a depth-2 meta-graph

$$\mathsf{Q}^{(2)} = (V^{(2)}, E^{(2)}, s^{(2)}, t^{(2)}, \lambda^{(2)}), \quad V^{(2)} = \{\mathsf{P}_1^{(1)}, \mathsf{P}_2^{(1)}\}, \quad E^{(2)} = \{r\},$$

with

$$s^{(2)}(r) = \mathsf{P}_1^{(1)}, \quad t^{(2)}(r) = \mathsf{P}_2^{(1)}, \quad \lambda^{(2)}(r) = (R_{\text{eq}}^\uparrow)^\uparrow.$$

To see $r \in (R_{\text{eq}}^\uparrow)^\uparrow$, take $X = \mathsf{M}_{\text{ester}} \in V(\mathsf{P}_1^{(1)})$ and $Y = \mathsf{M}_{\text{hydr}} \in V(\mathsf{P}_2^{(1)})$; we already have $(X, Y) \in R_{\text{eq}}^\uparrow$ (Example 6), so by the lift definition $(\mathsf{P}_1^{(1)}, \mathsf{P}_2^{(1)}) \in (R_{\text{eq}}^\uparrow)^\uparrow$. Thus $\mathsf{Q}^{(2)}$ is a Molecular Iterated MetaGraph of depth 2.

Theorem 2.19 (Molecular Iterated MetaGraphs are Iterated MetaGraphs). *For every $t \in \mathbb{N}_0$, any $\mathsf{M}^{(t)}$ as in Definition 2.14 is an iterated metagraph of depth t in the sense of the preliminaries (with base universe \mathfrak{Mol} and base relations $\mathcal{R}_{\text{chem}}$).*

Proof. By Definition 2.13, $(\mathfrak{Mol}^{(t)}, \mathcal{R}_{\text{chem}}^{(t)})$ is obtained from $(\mathfrak{Mol}^{(t-1)}, \mathcal{R}_{\text{chem}}^{(t-1)})$ by the same lifting scheme used for general iterated metagraphs. Thus $V^{(t)} \subseteq \mathfrak{Mol}^{(t)}$, $\lambda^{(t)}$ takes values in $\mathcal{R}_{\text{chem}}^{(t)}$, and each edge satisfies the required incidence. Therefore $\mathsf{M}^{(t)}$ is a metagraph over $(\mathfrak{Mol}^{(t)}, \mathcal{R}_{\text{chem}}^{(t)})$, i.e. an iterated metagraph of depth t . \square

3 Additional Results: Some Applications in Biochemistry

Biochemistry is the scientific study of chemical processes, reactions, and molecular interactions occurring within living organisms [21]. This section presents application examples of the Molecular MetaGraph and Molecular Iterated MetaGraph, as defined in this paper, within the field of biochemistry.

Example 3.1 (ATP-dependent phosphorylation (hexokinase step)). ATP-dependent phosphorylation is a biochemical process where ATP donates its phosphate group to a substrate, regulating energy transfer and cellular signaling (cf. [22, 23]). Let the molecular vertex set be

$$V = \{G_{\text{ATP}}, G_{\text{ADP}}, G_{\text{Glc}}, G_{\text{G6P}}\},$$

representing ATP, ADP, glucose, and glucose-6-phosphate. Model the hexokinase reaction



by the directed meta-edges

$$E = \{e_1, e_2\}, \quad s(e_1) = G_{\text{ATP}}, \quad t(e_1) = G_{\text{ADP}}, \quad \lambda(e_1) = R_{\text{rxn}}, \quad s(e_2) = G_{\text{Glc}}, \quad t(e_2) = G_{\text{G6P}}, \quad \lambda(e_2) = R_{\text{rxn}}.$$

Then

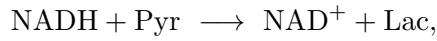
$$\mathsf{M}_{\text{HK}} := (V, E, s, t, \lambda)$$

is a Molecular MetaGraph over $(\mathfrak{Mol}, \mathcal{R}_{\text{chem}})$ because each edge satisfies the incidence constraint $(s(e_i), t(e_i)) \in R_{\text{rxn}} = \lambda(e_i)$ ($i = 1, 2$).

Example 3.2 (NADH-coupled reduction (lactate dehydrogenase step)). Let

$$V = \{G_{\text{NAD}^+}, G_{\text{NADH}}, G_{\text{Pyr}}, G_{\text{Lac}}\},$$

representing NAD^+ , NADH, pyruvate, and lactate. For the LDH reaction



take

$$E = \{e_1, e_2\}, \quad s(e_1) = G_{\text{NADH}}, t(e_1) = G_{\text{NAD}^+}, \lambda(e_1) = R_{\text{rxn}}, \quad s(e_2) = G_{\text{Pyr}}, t(e_2) = G_{\text{Lac}}, \lambda(e_2) = R_{\text{rxn}}.$$

Thus $\mathbf{M}_{\text{LDH}} = (V, E, s, t, \lambda)$ is a valid Molecular MetaGraph since $(s(e_i), t(e_i)) \in R_{\text{rxn}} = \lambda(e_i)$ for $i = 1, 2$.

We use the lifted relations of Definition 2.13. Recall:

$$(\mathbf{M}_1, \mathbf{M}_2) \in R^\uparrow \iff \exists X \in V(\mathbf{M}_1), \exists Y \in V(\mathbf{M}_2) \text{ with } (X, Y) \in R.$$

Example 3.3 (Depth $t = 1$: Linking consecutive glycolytic steps via a lifted equality). Let \mathbf{M}_{HK} be the hexokinase meta-graph from the first example. Define the phosphoglucose isomerase (PGI) [24] meta-graph

$$\mathbf{M}_{\text{PGI}} : V' = \{G_{\text{G6P}}, G_{\text{F6P}}\}, \quad E' = \{e'_1, e'_2\},$$

with

$$s(e'_1) = G_{\text{G6P}}, t(e'_1) = G_{\text{F6P}}, \lambda(e'_1) = R_{\text{rxn}}, \quad s(e'_2) = G_{\text{F6P}}, t(e'_2) = G_{\text{G6P}}, \lambda(e'_2) = R_{\text{rxn}}.$$

Form a depth-1 iterated meta-graph

$$\mathbf{N}_{\text{Gly}}^{(1)} = (V^{(1)}, E^{(1)}, s^{(1)}, t^{(1)}, \lambda^{(1)}),$$

with

$$V^{(1)} = \{\mathbf{M}_{\text{HK}}, \mathbf{M}_{\text{PGI}}\}, \quad E^{(1)} = \{f\}, \quad s^{(1)}(f) = \mathbf{M}_{\text{HK}}, \quad t^{(1)}(f) = \mathbf{M}_{\text{PGI}}, \quad \lambda^{(1)}(f) = R_{\text{eq}}^\uparrow.$$

Since $G_{\text{G6P}} \in V(\mathbf{M}_{\text{HK}}) \cap V(\mathbf{M}_{\text{PGI}})$, we have $(G_{\text{G6P}}, G_{\text{G6P}}) \in R_{\text{eq}}$, so by definition $(s^{(1)}(f), t^{(1)}(f)) \in R_{\text{eq}}^\uparrow = \lambda^{(1)}(f)$. Hence $\mathbf{N}_{\text{Gly}}^{(1)}$ is a valid Molecular Iterated MetaGraph of depth 1.

Example 3.4 (Depth $t = 2$: Dehydrogenase family coupling via double lift). (cf. [25, 26]) Construct three Molecular MetaGraphs on redox-coupled steps:

- (i) LDH (already defined) with molecules $\{G_{\text{NAD}^+}, G_{\text{NADH}}, G_{\text{Pyr}}, G_{\text{Lac}}\}$.
- (ii) MDH (malate dehydrogenase):

$$\mathbf{M}_{\text{MDH}} : V = \{G_{\text{NAD}^+}, G_{\text{NADH}}, G_{\text{OAA}}, G_{\text{Mal}}\}, \quad E = \{e_1, e_2\},$$

$$s(e_1) = G_{\text{NADH}}, t(e_1) = G_{\text{NAD}^+}, \lambda(e_1) = R_{\text{rxn}}, \quad s(e_2) = G_{\text{OAA}}, t(e_2) = G_{\text{Mal}}, \lambda(e_2) = R_{\text{rxn}}.$$

- (iii) GAPDH (glyceraldehyde-3-phosphate dehydrogenase):

$$\mathbf{M}_{\text{GAPDH}} : V = \{G_{\text{NAD}^+}, G_{\text{NADH}}, G_{\text{G3P}}, G_{1,3\text{-BPG}}\}, \quad E = \{e_1, e_2\},$$

$$s(e_1) = G_{\text{NAD}^+}, t(e_1) = G_{\text{NADH}}, \lambda(e_1) = R_{\text{rxn}}, \quad s(e_2) = G_{\text{G3P}}, t(e_2) = G_{1,3\text{-BPG}}, \lambda(e_2) = R_{\text{rxn}}.$$

Create two depth-1 nodes using lifted equality (they share NAD^+/NADH):

$$\mathbf{P}_1^{(1)} = (\{\mathbf{M}_{\text{LDH}}, \mathbf{M}_{\text{MDH}}\}, \{p\}, s^{(1)}, t^{(1)}, \lambda^{(1)})$$

with $s^{(1)}(p) = M_{LDH}$, $t^{(1)}(p) = M_{MDH}$, $\lambda^{(1)}(p) = R_{eq}^\uparrow$, witnessed by the common molecule G_{NAD^+} (or G_{NADH}); and

$$P_2^{(1)} = (\{M_{GAPDH}, M_{LDH}\}, \{q\}, s^{(1)}, t^{(1)}, \lambda^{(1)})$$

with $s^{(1)}(q) = M_{GAPDH}$, $t^{(1)}(q) = M_{LDH}$, $\lambda^{(1)}(q) = R_{eq}^\uparrow$, witnessed again by $NAD^+/NADH$.

Now define a depth-2 Molecular Iterated MetaGraph

$$Q^{(2)} = (V^{(2)}, E^{(2)}, s^{(2)}, t^{(2)}, \lambda^{(2)}), \quad V^{(2)} = \{P_1^{(1)}, P_2^{(1)}\}, \quad E^{(2)} = \{r\},$$

with

$$s^{(2)}(r) = P_1^{(1)}, \quad t^{(2)}(r) = P_2^{(1)}, \quad \lambda^{(2)}(r) = (R_{eq}^\uparrow)^\uparrow.$$

To verify incidence, choose $X = M_{LDH} \in V(P_1^{(1)})$, $Y = M_{LDH} \in V(P_2^{(1)})$. Because X and Y each contain G_{NAD^+} (hence $(G_{NAD^+}, G_{NAD^+}) \in R_{eq}$), we get $(X, Y) \in R_{eq}^\uparrow$, which witnesses $(s^{(2)}(r), t^{(2)}(r)) \in (R_{eq}^\uparrow)^\uparrow$. Therefore $Q^{(2)}$ is a valid depth-2 Molecular Iterated MetaGraph.

4 Conclusion

In this paper, we studied new extensions called the *Molecular MetaGraph* and the *Molecular Iterated MetaGraph*, which generalize the notion of molecular graphs using the frameworks of MetaGraphs and Iterated MetaGraphs.

In future work, we aim to investigate further extensions based on alternative graph frameworks, such as *Fuzzy Graphs* [27], *Neutrosophic Graphs* [28,29], *Directed Graphs* [30], *HyperFuzzy Graphs* [31], *HyperGraph* [32], *SuperHyperGraph* [33,34], *Bidirected Graphs* [35], and *Plithogenic Graphs* [36]. Moreover, we hope that this line of research will advance toward applications in the field of chemistry as well as the development of algorithms.

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Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this work.

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Data Availability

This paper is theoretical and did not generate or analyze any empirical data. We welcome future studies that apply and test these concepts in practical settings.

Research Integrity

The author confirms that this manuscript is original, has not been published elsewhere, and is not under consideration by any other journal.

Use of Computational Tools

All proofs and derivations were performed manually; no computational software (e.g., Mathematica, Sage-Math, Coq) was used.

Code Availability

No code or software was developed for this study.

Ethical Approval

This research did not involve human participants or animals, and therefore did not require ethical approval.

Use of Generative AI and AI-Assisted Tools

We use generative AI and AI-assisted tools for tasks such as English grammar checking, and We do not employ them in any way that violates ethical standards.

Supplementary Information

No supplementary materials accompany this paper.

Disclaimer

The ideas presented here are theoretical and have not yet been validated through empirical testing. While we have strived for accuracy and proper citation, inadvertent errors may remain. Readers should verify any referenced material independently. The opinions expressed are those of the authors and do not necessarily reflect the views of their institutions.

References

- [1] Reinhard Diestel. *Graph theory*. Springer (print edition); Reinhard Diestel (eBooks), 2024.
- [2] Jonathan L Gross, Jay Yellen, and Mark Anderson. *Graph theory and its applications*. Chapman and Hall/CRC, 2018.
- [3] RB Azevedo, Rolf Lohaus, and Tiago Paixão. Networking networks. *Evol Dev*, 10:514–515, 2008.
- [4] Claire Donnat and Susan Holmes. Tracking network dynamics: A survey using graph distances. *The Annals of Applied Statistics*, 12(2):971–1012, 2018.
- [5] Olivier C Martin and Andreas Wagner. Multifunctionality and robustness trade-offs in model genetic circuits. *Biophysical journal*, 94(8):2927–2937, 2008.
- [6] Jiaqi Cao, Shengli Zhang, Qingxia Chen, Houtian Wang, Mingzhe Wang, and Naijin Liu. Network-wide task offloading with leo satellites: A computation and transmission fusion approach. *arXiv preprint arXiv:2211.09672*, 2022.

- [7] Takaaki Fujita. Metahypergraphs, metasuperhypergraphs, and iterated metagraphs: Modeling graphs of graphs, hypergraphs of hypergraphs, superhypergraphs of superhypergraphs, and beyond. *Current Research in Interdisciplinary Studies*, 4(5):1–23, 2025.
- [8] Steven Kearnes, Kevin McCloskey, Marc Berndl, Vijay Pande, and Patrick Riley. Molecular graph convolutions: moving beyond fingerprints. *Journal of computer-aided molecular design*, 30:595–608, 2016.
- [9] Ivan Gutman and Ernesto Estrada. Topological indices based on the line graph of the molecular graph. *Journal of chemical information and computer sciences*, 36(3):541–543, 1996.
- [10] Jiaxuan You, Bowen Liu, Zhitao Ying, Vijay Pande, and Jure Leskovec. Graph convolutional policy network for goal-directed molecular graph generation. *Advances in neural information processing systems*, 31, 2018.
- [11] Hiroshi Kajino. Molecular hypergraph grammar with its application to molecular optimization. In *International Conference on Machine Learning*, pages 3183–3191. PMLR, 2019.
- [12] Elena V Konstantinova and Vladimir A Skorobogatov. Molecular hypergraphs: The new representation of nonclassical molecular structures with polycentric delocalized bonds. *Journal of chemical information and computer sciences*, 35(3):472–478, 1995.
- [13] Takaaki Fujita. Note for biochemical/electrochemical/physicochemical hypergraphs and superhypergraphs. *Information Sciences and Technological Innovations*, 2025.
- [14] Takaaki Fujita. An introduction and reexamination of molecular hypergraph and molecular n-superhypergraph. *Asian Journal of Physical and Chemical Sciences*, 13(3):1–38, 2025.
- [15] Ramón García-Domenech, Jorge Gálvez, Jesus V de Julián-Ortiz, and Lionello Pogliani. Some new trends in chemical graph theory. *Chemical Reviews*, 108(3):1127–1169, 2008.
- [16] Stephan Wagner and Hua Wang. *Introduction to chemical graph theory*. Chapman and Hall/CRC, 2018.
- [17] Takaaki Fujita and Florentin Smarandache. *A reconsideration of advanced concepts in neutrosophic graphs: Smart, zero divisor, layered, weak, semi, and chemical graphs*. Infinite Study, 2024.
- [18] Takaaki Fujita. Molecular fuzzy graphs, hypergraphs, and superhypergraphs. *Journal of Intelligent Decision and Computational Modelling*, 1(3):158–171, 2025.
- [19] Johannes Gasteiger, Janek Groß, and Stephan Günemann. Directional message passing for molecular graphs. *arXiv preprint arXiv:2003.03123*, 2020.
- [20] Wengong Jin, Regina Barzilay, and Tommi Jaakkola. Hierarchical generation of molecular graphs using structural motifs. In *International conference on machine learning*, pages 4839–4848. PMLR, 2020.
- [21] Mary K Campbell and Sharm O Farrell. *Biochemistry*. 2009.
- [22] Jesse C Hay, Phillip L Fisette, Glenn H Jenkins, Kiyoko Fukami, Tadaomi Takenawa, Richard A Anderson, and Thomas FJ Martin. Atp-dependent inositol phosphorylation required for ca2+-activated secretion. *Nature*, 374(6518):173–177, 1995.
- [23] Jing Jing Ye, Jonathan Reizer, Xuewen Cui, and Milton H Saier Jr. Atp-dependent phosphorylation of serine-46 in the phosphocarrier protein hpr regulates lactose/h+ symport in lactobacillus brevis. *Proceedings of the National Academy of Sciences*, 91(8):3102–3106, 1994.
- [24] T Yanagawa, T Funasaka, S Tsutsumi, H Watanabe, and A Raz. Novel roles of the autocrine motility factor/phosphoglucose isomerase in tumor malignancy. *Endocrine-Related Cancer*, 11(4):749–759, 2004.
- [25] James G Ferry. Co dehydrogenase. *Annual Review of Microbiology*, 49:305–334, 1995.
- [26] Christopher Anthony and Paul Williams. The structure and mechanism of methanol dehydrogenase. *Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics*, 1647(1-2):18–23, 2003.
- [27] Azriel Rosenfeld. Fuzzy graphs. In *Fuzzy sets and their applications to cognitive and decision processes*, pages 77–95. Elsevier, 1975.
- [28] S. N. Suber Bathusha, Sowndharya Jayakumar, S. Angelin, and Kavitha Raj. The energy of interval-valued complex neutrosophic graph structures: Framework, application and future research directions. *Neutrosophic Systems with Applications*, 2024.
- [29] Said Broumi, Mohamed Talea, Assia Bakali, and Florentin Smarandache. Single valued neutrosophic graphs. *Journal of New theory*, 10:86–101, 2016.
- [30] Jinta Jose, Bobin George, and Rajesh K Thumabakara. Soft directed graphs, their vertex degrees, associated matrices and some product operations. *New Mathematics and Natural Computation*, 19(03):651–686, 2023.
- [31] Takaaki Fujita. *Advancing Uncertain Combinatorics through Graphization, Hyperization, and Uncertainization: Fuzzy, Neutrosophic, Soft, Rough, and Beyond*. Biblio Publishing, 2025.

- [32] Claude Berge. *Hypergraphs: combinatorics of finite sets*, volume 45. Elsevier, 1984.
- [33] Masoud Ghods, Zahra Rostami, and Florentin Smarandache. Introduction to neutrosophic restricted superhypergraphs and neutrosophic restricted superhypertrees and several of their properties. *Neutrosophic Sets and Systems*, 50:480–487, 2022.
- [34] Florentin Smarandache. *Extension of HyperGraph to n-SuperHyperGraph and to Plithogenic n-SuperHyperGraph, and Extension of HyperAlgebra to n-ary (Classical-/Neutro-/Anti-) HyperAlgebra*. Infinite Study, 2020.
- [35] Soumalya Joardar and Atibur Rahaman. Almost complex structure on finite points from bidirected graphs. *Journal of Noncommutative Geometry*, 2023.
- [36] Fazeelat Sultana, Muhammad Gulistan, Mumtaz Ali, Naveed Yaqoob, Muhammad Khan, Tabasam Rashid, and Tauseef Ahmed. A study of plithogenic graphs: applications in spreading coronavirus disease (covid-19) globally. *Journal of ambient intelligence and humanized computing*, 14(10):13139–13159, 2023.